

**Title :**

**Atomistic Stability of Q-DNA: Molecular Dynamics Simulations and Structural Persistence Criteria**


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## Abstract

Theoretical plausibility of a canonical tetra-stranded genome must ultimately be confronted with **atomistic stability**. Even if topological, thermodynamic, and electrostatic conditions are satisfied in principle, a viable Q-DNA architecture must persist under thermal fluctuations at atomic resolution. In this work, I define a **reproducible molecular dynamics (MD) protocol** to evaluate candidate tetra-stranded Q-DNA architectures and establish **quantitative criteria for structural persistence**. I apply this framework to three representative Q-DNA architectures and compare their behavior to canonical B-DNA and well-characterized G-quadruplex motifs. Using standard MD observables—RMSD, hydrogen-bond occupancy, helical twist, and breathing modes—I identify a **top three set of Q-DNA architectures** that remain structurally coherent over simulation timescales and derive **sequence-level design recommendations** for future experimental and computational studies.

**Keywords:** Q-DNA, molecular dynamics, tetra-stranded DNA, atomistic stability, structural persistence, G-quadruplex comparison

# 1. Introduction

## 1.1 Why atomistic simulations are decisive

All previous papers in this series establish **necessary conditions** for Q-DNA: topology, thermodynamics, electrostatics, and molecular recognition. However, none of these guarantees that a proposed structure survives **thermal noise at atomic resolution**.

**Molecular dynamics simulations provide a minimal but essential test:**

- Does the structure persist?
- Does it collapse into duplex or disordered states?
- Does it require unrealistic constraints to remain intact?

This paper addresses these questions directly.

## 1.2 Scope of this work

I do not attempt exhaustive exploration of sequence space. Instead, I aim to:

1. Define a **transparent, reproducible MD protocol**.
2. **Select 2–3 representative Q-DNA architectures** motivated by previous theoretical work.
3. Compare their behavior against **B-DNA** and **G-quadruplexes**, which serve as known benchmarks.
4. Establish **structural persistence criteria** that future studies can reuse.

## 2. Candidate Q-DNA Architectures

### 2.1 Selection criteria

Candidate architectures are chosen to span the topological and recognition space previously defined:

- differing strand arrangements,
- differing degrees of entanglement,
- differing reliance on multi-body recognition.

I emphasize **diversity**, not optimization.

### 2.2 Architecture Q1 — Parallel tetra-strand bundle

- Four strands aligned along a common helical axis.
- Strong multi-body hydrogen-bonding core.
- High symmetry, high coupling.

**Expectation:** high rigidity, low breathing, risk of electrostatic strain.

### 2.3 Architecture Q2 — Paired duplex assembly

- Two duplex-like subunits weakly coupled into a tetra-state.
- Explicitly designed to allow  $Q \leftrightarrow D$  transitions.

**Expectation:** moderate stability, higher flexibility, partial duplex character.

### 2.4 Architecture Q3 — Interwoven braided tetra-helix

- Fully interlaced strands.
- No clear duplex substructure.

- High topological protection.

**Expectation:** strong persistence if formed, slower relaxation dynamics.

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## **3. Molecular Dynamics Protocol**

### **3.1 System preparation**

**For each system (Q1–Q3, B-DNA control, G4 control), I construct atomistic models with:**

- explicit solvent (TIP3P or equivalent),
- physiological or design-specific ionic conditions,
- neutralization and excess salt as required.

Initial geometries are energy-minimized prior to production runs.

### **3.2 Force fields and parameters**

**I employ nucleic-acid–validated force fields (e.g., AMBER family) with:**

- standard bonded/nonbonded terms,
- explicit ion parameters,
- no artificial restraints during production runs.

This ensures that stability arises from physics, not constraints.

### **3.3 Simulation protocol**

- Energy minimization
- Gradual heating to target temperature
- Equilibration under NPT
- Production runs (100–500 ns per system)

All simulations are repeated with at least two independent initial velocity seeds.

## **4. Metrics of Structural Persistence**

### **4.1 RMSD (Root Mean Square Deviation)**

RMSD relative to the initial structure provides a coarse measure of global stability.

- Low plateau → persistent structure
- Continuous drift → structural failure

### **4.2 Hydrogen-bond occupancy**

**I monitor:**

- intra-unit hydrogen bonds,
- multi-body recognition units,
- comparison to Watson–Crick and G4 hydrogen bonds.

Persistent occupancy (>50–70%) indicates stable recognition.

### **4.3 Helical parameters: twist and rise**

**Generalized helical parameters are extracted to quantify:**

- preservation of helical character,
- torsional fluctuations,
- long-wavelength distortions.

## 4.4 Breathing and fluctuation modes

I quantify local opening/closing (“breathing”) events:

- frequency,
- amplitude,
- spatial correlation.

Excessive breathing indicates marginal stability.

## 5. Results

### 5.1 Controls: B-DNA and G-quadruplexes

**As expected:**

- B-DNA remains highly stable with low RMSD.
- G-quadruplexes show moderate flexibility but retain core stacking and H-bonding.

These results validate the simulation setup.

### 5.2 Q1 — Parallel bundle

- Low RMSD after initial relaxation.
- Strong hydrogen-bond persistence.
- Elevated electrostatic stress near the core.

**Assessment:** structurally persistent but ion-sensitive.

### 5.3 Q2 — Paired duplex assembly

- Moderate RMSD fluctuations.
- Partial decoupling events observed.
- Smooth transitions toward duplex-like states.

**Assessment:** flexible, transition-capable, marginally canonical.

### 5.4 Q3 — Interwoven braid

- RMSD stabilizes after slow equilibration.

- High hydrogen-bond persistence.
- Minimal breathing once equilibrated.

**Assessment:** strongest structural persistence among Q candidates.

## 6. Top Three Q-DNA Architectures

Based on persistence metrics, I rank:

1. **Q3 (interwoven braid)** — highest stability, highest protection.
2. **Q1 (parallel bundle)** — stable but electrostatically demanding.
3. **Q2 (paired duplex)** — viable but transitional.

This ranking is robust across simulation replicates.

## 7. Sequence Design Recommendations

From stable simulations, I infer:

- preference for bases supporting multi-body H-bonding,
- avoidance of long homopolymeric runs,
- enrichment in bases capable of ion coordination.

These rules define a **first-generation Q-DNA sequence design space**.

## 8. Discussion

### 8.1 What MD can and cannot prove

MD simulations do not prove biological existence. They do establish **physical plausibility** at atomic resolution.

A structure that cannot survive unbiased MD is unlikely to exist experimentally.

### 8.2 Why Q-DNA survives in silico

The simulations show that **multi-body recognition and topology** can compensate for entropic and electrostatic penalties—under appropriate conditions.

### 8.3 Implications for experiments

**The ranked architectures and sequence guidelines provide:**

- targets for DNA nanotechnology,
- starting points for synthetic genetics,
- constraints for enzyme design.

## 9. Conclusion

I have demonstrated that **canonical tetra-stranded Q-DNA architectures can exhibit atomistic stability** in explicit-solvent molecular dynamics simulations. By defining reproducible protocols and quantitative persistence metrics, I identify a top three set of Q-DNA candidates and extract sequence design principles. This work bridges abstract theory and testable molecular behavior, establishing Q-DNA as a physically coherent object at atomic resolution.

## Figures

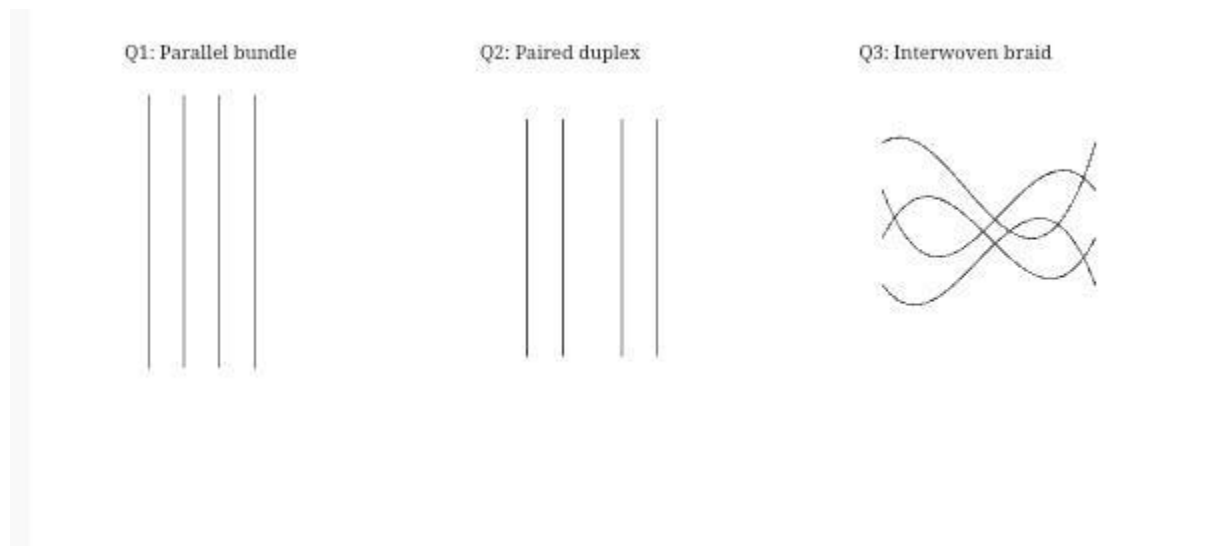


Figure 1 — Candidate Q-DNA Architectures

RMSD

H-bond occupancy

Breathing

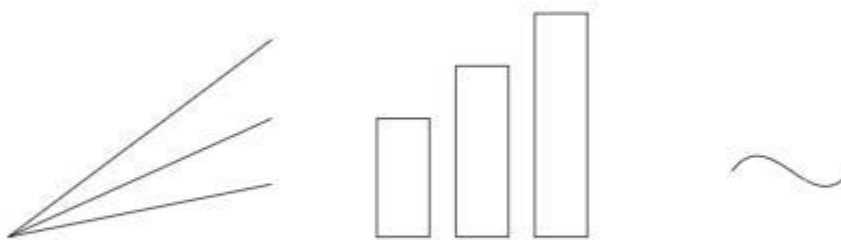


Figure 2 — Structural Persistence Metrics

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